FILE 'CAPLUS, MEDLINE, BIOSIS, EMBASE' ENTERED AT 13:02:57 ON 31 MAR 2004

L1	1090	S	JIANG.IN.
L2	8	S	L1 AND FUSION PROTEIN
L3	2	DUP	REM L2 (6 DUPLICATES REMOVED)
L4	0	S	EXPRES### SAME FUSION PROTEIN SAME REPOTER SAME LIBRAR###
L5	35	S	EXPRES###(10A) FUSION PROTEIN (10A) REPORTER#
L6	4	S	L5 AND LIBRAR###
L7	0	S	L6 AND CYCLOHEXIMIDE
L8	4	DUP	REM L6 (0 DUPLICATES REMOVED)
L9	0	S	FUSION(10A)GREEN FLUORESCENCE(10A)LIBRAR###
L10	40	S	FUSION(10A)GREEN(10A)PROTEIN(10A)LIBRAR###
L11	15	S	L10 AND EXPRES###
L12	0	S	L11 AND CYCLOHEXIMIDE
L13	10	S	L11 AND SCREEN###
L14	0	S	L13 AND (INHIBIT###(10A)DEGRADAT###)
L15	0	S	L13 AND INHIBIT###
L16	9	DUP	REM L13 (1 DUPLICATE REMOVED)
L17	223458	S	LI.IN.
L18	153	S	L17 AND CYCLOHEXIMIDE
L19	0	S	L18 AND FLUORESCENT PROTEIN
L20	0	S	L18 AND FUSION PROTEIN
L21	37	S	L17 AND FLUORESCENT PROTEIN
L22	0	S	L21 AND LIBRAR###
L23	1	S	L21 AND SCREEN###

10053516

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FILE 'MEDLINE' ENTERED AT 13:02:57 ON 31 MAR 2004

FILE 'BIOSIS' ENTERED AT 13:02:57 ON 31 MAR 2004
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FILE 'EMBASE' ENTERED AT 13:02:57 ON 31 MAR 2004
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=> s Jiang.in.
L1 1090 JIANG.IN.

=> s l1 and fusion protein
L2 8 L1 AND FUSION PROTEIN

=> dup rem l12
L12 IS NOT VALID HERE
The L-number entered has not been defined in this session, or it has been deleted. To see the L-numbers currently defined in this session, enter DISPLAY HISTORY at an arrow prompt (=>).

=> dup rem l2
PROCESSING COMPLETED FOR L2
L3 2 DUP REM L2 (6 DUPLICATES REMOVED)

=> d l3 1-2 bib ab kwic

L3 ANSWER 1 OF 2 CAPLUS COPYRIGHT 2004 ACS on STN DUPLICATE 1
AN 1999:320171 CAPLUS
DN 131:100245
TI Expression of a truncated receptor protein tyrosine phosphatase kappa in the brain of an adult transgenic mouse
AU Shen, P.; Canoll, P. D.; Sap, J.; Musacchio, J. M.
CS Department of Pharmacology, New York University Medical Center, New York, NY, 10016, USA
SO Brain Research (1999), 826(2), 157-171
CODEN: BRREAP; ISSN: 0006-8993
PB Elsevier Science B.V.
DT Journal
LA English
AB Receptor protein tyrosine phosphatases (RPTPs) comprise a family of proteins that feature intracellular phosphatase domains and an ectodomain with putative ligand-binding motifs. Several RPTPs are expressed in the brain, including RPTP-K which participates in homophilic cell-cell interactions in vitro (**Jiang**, Y.-P et al., 1993; Sap, J. et al., 1994). The homol. of RPTP-κ's ectodomain to neural cell adhesion mols. indicates potential roles in developmental processes such as axonal growth and target recognition, as has been demonstrated for certain Drosophila RPTPs. The brain distribution of RPTP-κ-expressing cells has not been determined, however. In a gene-trap mouse model with a β-gal + neo (1-geo) insertion in the endogenous RPTP-κ gene, the consequent loss of RPTP-κ's enzymic activity does not produce any obvious phenotypic defects (Skarnes, W. C. et al., 1995). Nevertheless, since the transgene's expression is driven by the endogenous RPTP-κ promoter, distribution of the truncated RPTP-κ/β-geo **fusion protein** should reflect the regional and cellular expression of wild-type RPTP-κ, and thus may identify sites where RPTP-κ is important. Towards that goal, the authors have used this mouse model to map the distribution of the truncated RPTP-κ/β-geo **fusion protein** in the adult mouse brain using

β-galactosidase as a marker enzyme. Visualization of the β-galactosidase activity revealed a non-random pattern of expression, and identified cells throughout the CNS that display RPTP-κ promoter activity. Several neural systems highly expressed the transgene-most notably cortical, olfactory, hippocampal, hypothalamic, amygdaloid and visual structures. These well-characterized brain regions may provide a basis for future studies of RPTP-κ function.

RE.CNT 40 THERE ARE 40 CITED REFERENCES AVAILABLE FOR THIS RECORD
ALL CITATIONS AVAILABLE IN THE RE FORMAT

AB Receptor protein tyrosine phosphatases (RPTPs) comprise a family of proteins that feature intracellular phosphatase domains and an ectodomain with putative ligand-binding motifs. Several RPTPs are expressed in the brain, including RPTP-K which participates in homophilic cell-cell interactions in vitro (**Jiang**, Y.-P et al., 1993; Sap, J. et al., 1994). The homol. of RPTP-κ's ectodomain to neural cell adhesion mols. indicates potential roles in developmental processes such as axonal growth and target recognition, as has been demonstrated for certain Drosophila RPTPs. The brain distribution of RPTP-κ-expressing cells has not been determined, however. In a gene-trap mouse model with a β-gal + neo (1-geo) insertion in the endogenous RPTP-κ gene, the consequent loss of RPTP-κ's enzymic activity does not produce any obvious phenotypic defects (Skarnes, W. C. et al., 1995). Nevertheless, since the transgene's expression is driven by the endogenous RPTP-κ promoter, distribution of the truncated RPTP-κ/β-geo **fusion protein** should reflect the regional and cellular expression of wild-type RPTP-κ, and thus may identify sites where RPTP-κ is important. Towards that goal, the authors have used this mouse model to map the distribution of the truncated RPTP-κ/β-geo **fusion protein** in the adult mouse brain using β-galactosidase as a marker enzyme. Visualization of the β-galactosidase activity revealed a non-random pattern of expression, and identified cells throughout the CNS that display RPTP-κ promoter activity. Several neural systems highly expressed the transgene-most notably cortical, olfactory, hippocampal, hypothalamic, amygdaloid and visual structures. These well-characterized brain regions may provide a basis for future studies of RPTP-κ function.

L3 ANSWER 2 OF 2 CAPLUS COPYRIGHT 2004 ACS on STN DUPLICATE 2
AN 1997:698880 CAPLUS
DN 128:31575
TI Cloning and expression of a gene encoding a protein obtained from earthworm secretion that is a chemoattractant for garter snakes
AU Liu, Weimin; Wang, Dalton; Chen, Ping; Halpern, Mimi
CS Department of Biochemistry, SUNY Health Science Center at Brooklyn, NY, 11203, USA
SO Journal of Biological Chemistry (1997), 272(43), 27378-27381
CODEN: JBCHA3; ISSN: 0021-9258
PB American Society for Biochemistry and Molecular Biology
DT Journal
LA English
AB The protein ES20, derived from earthworm shock secretion, is a vomeronasally mediated chemoattractant for garter snakes (**Jiang**, X. C., Inouchi, J., Wang, D., and Halpern, M. (1990) J. Biol. Chemical 265, 8736-8744). Based on its 15-residue N-terminal amino acid sequence, degenerative oligodeoxynucleotide probes were synthesized and used to screen a cDNA library that was constructed in sense orientation using a Uni-ZAP XR vector and XL1-Blue MRF' host. A gene was cloned from a polymerase chain reaction as well as from the cDNA library. A combination of the forward degenerative primer and T7 primer was used to obtain gene-specific DNA fragments, from which probes were synthesized and successfully used in screening the cDNA library. The ES20 gene is about 700 base pairs long and encodes 208 amino residues. The ES20 gene was excised from a recombinant plasmid pSK-ES20, ligated to pQE30 expression vector, and transformed into Escherichia coli strain JM109. The selected

recombinant plasmids were transformed into expression host cell, E. coli M15[pREP4]. Three transformants were selected, induced with isopropyl-1-thio-β-D-galactopyranoside for fusion gene expression and an expressed 20-kDa **fusion protein** purified under denaturing conditions. This protein was refolded and gave a pos. reaction against ES20-specific polyclonal antibodies. The **fusion protein** that had not been denatured remained as an aggregate and was an active chemoattractant for garter snakes.

RE.CNT 19 THERE ARE 19 CITED REFERENCES AVAILABLE FOR THIS RECORD
ALL CITATIONS AVAILABLE IN THE RE FORMAT

AB The protein ES20, derived from earthworm shock secretion, is a vomeronasally mediated chemoattractant for garter snakes (**Jiang**, X. C., Inouchi, J., Wang, D., and Halpern, M. (1990) J. Biol. Chemical 265, 8736-8744). Based on its 15-residue N-terminal amino acid sequence, degenerative oligodeoxynucleotide probes were synthesized and used to screen a cDNA library that was constructed in sense orientation using a Uni-ZAP XR vector and XL1-Blue MRF' host. A gene was cloned from a polymerase chain reaction as well as from the cDNA library. A combination of the forward degenerative primer and T7 primer was used to obtain gene-specific DNA fragments, from which probes were synthesized and successfully used in screening the cDNA library. The ES20 gene is about 700 base pairs long and encodes 208 amino residues. The ES20 gene was excised from a recombinant plasmid pSK-ES20, ligated to pQE30 expression vector, and transformed into Escherichia coli strain JM109. The selected recombinant plasmids were transformed into expression host cell, E. coli M15[pREP4]. Three transformants were selected, induced with isopropyl-1-thio-β-D-galactopyranoside for fusion gene expression and an expressed 20-kDa **fusion protein** purified under denaturing conditions. This protein was refolded and gave a pos. reaction against ES20-specific polyclonal antibodies. The **fusion protein** that had not been denatured remained as an aggregate and was an active chemoattractant for garter snakes.

```
=> s expres### same fusion protein same repoter same librar###
L4            0 EXPRES### SAME FUSION PROTEIN SAME REPOTER SAME LIBRAR###

=> s expres###(10a) fusion protein (10a) reporter#
L5           35 EXPRES###(10A) FUSION PROTEIN (10A) REPORTER#

=> s l5 and librar###
L6            4 L5 AND LIBRAR###

=> s l6 and cycloheximide
L7            0 L6 AND CYCLOHEXIMIDE

=> dup rem l6
PROCESSING COMPLETED FOR L6
L8            4 DUP REM L6 (0 DUPLICATES REMOVED)

=> d l8 1-4 bib ab kwic

L8   ANSWER 1 OF 4  CAPLUS  COPYRIGHT 2004 ACS on STN
AN   2003:656310  CAPLUS
DN   139:174827
TI   Methods and kits for isolating and characterizing short-lived proteins and
     arrays derived therefrom for use in drug screening
IN   Li, Xianqiang; Jiang, Xin
PA   USA
SO   U.S. Pat. Appl. Publ., 30 pp., Cont.-in-part of U.S. Ser. No. 53,230.
     CODEN: USXXCO
DT   Patent
LA   English
FAN.CNT 3
```

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	US 2003157540	A1	20030821	US 2003-347160	20030116
	US 2003134287	A1	20030717	US 2002-53230	20020116
	US 2003134288	A1	20030717	US 2002-53516	20020116
PRAI	US 2002-53230	A2	20020116		
	US 2002-53516	A2	20020116		

AB Compns., kits and methods are provided for isolating and characterizing short-lived proteins. The method comprises taking a **library** of cells wherein each cell in the **library expresses** a **fusion protein** comprising a **reporter** protein and a protein encoded by a sequence from a cDNA **library** derived from a sample of cells.. The sequence from the cDNA **library** is varied within the cell **library** and the rate of protein expression or degradation by cells in the **library** is modified. A population of cells is selected from the **library** of cells based on the population of cells having different reporter signal intensities than other cells in the **library** wherein the difference between intensities is indicative of the population of cells expressing shorter lived fusion proteins than the fusion proteins expressed by the other cells in the **library** and determining protein sequences of the fusion proteins of the selected population of cells. Also provided are oligonucleotide, protein and antibody arrays derived from short-lived proteins. The arrays can be used for efficiently profiling expression of short-lived proteins, screening for binding agents and comparing expression levels under different conditions.

AB Compns., kits and methods are provided for isolating and characterizing short-lived proteins. The method comprises taking a **library** of cells wherein each cell in the **library expresses** a **fusion protein** comprising a **reporter** protein and a protein encoded by a sequence from a cDNA **library** derived from a sample of cells.. The sequence from the cDNA **library** is varied within the cell **library** and the rate of protein expression or degradation by cells in the **library** is modified. A population of cells is selected from the **library** of cells based on the population of cells having different reporter signal intensities than other cells in the **library** wherein the difference between intensities is indicative of the population of cells expressing shorter lived fusion proteins than the fusion proteins expressed by the other cells in the **library** and determining protein sequences of the fusion proteins of the selected population of cells. Also provided are oligonucleotide, protein and antibody arrays derived from short-lived proteins. The arrays can be used for efficiently profiling expression of short-lived proteins, screening for binding agents and comparing expression levels under different conditions.

IT Animal cell
(**library**; methods and kits for isolating and characterizing short-lived proteins and arrays derived therefrom for use in drug screening)

IT cDNA **library**
(proteins encoded by; methods and kits for isolating and characterizing short-lived proteins and arrays derived therefrom for use in drug screening)

L8 ANSWER 2 OF 4 CAPLUS COPYRIGHT 2004 ACS on STN
AN 2002:353656 CAPLUS
DN 136:351377
TI Screening genes encoding nuclear transport proteins by GFP fusion protein expression and cell sorting
IN Maekawa, Takami; Mori, Maiko; Takahara, Yoshiyuki
PA Ajinomoto Co.,Inc., Japan
SO PCT Int. Appl., 60 pp.
CODEN: PIXXD2
DT Patent

LA Japanese
FAN.CNT 1

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	WO 2002036823	A1	20020510	WO 2001-JP9700	20011106
	W: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM				
	RW: GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR, BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG				
	AU 2002011021	A5	20020515	AU 2002-11021	20011106
	EP 1333099	A1	20030806	EP 2001-979034	20011106
	R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, SI, LT, LV, FI, RO, MK, CY, AL, TR				
	US 2003180801	A1	20030925	US 2002-204310	20020821
PRAI	JP 2000-337906	A	20001106		
	WO 2001-JP9700	W	20011106		

AB A method for screening a gene encoding a protein specifically transporting into nucleus by cloning each gene in a group of a large number of genes into an expression vector to **express** as **fusion protein** with a marker or **reporter** protein to construct a gene expression **library**, introducing this gene expression **library** into two cell groups to express the fusion proteins, stimulating one of the cell groups, separating cells wherein the fusion proteins are localized in the nucleus of such cells from both cell groups, and comparing the separated cells or nuclei to thereby search for a gene encoding a protein specifically transporting into the nucleus upon stimulus, is disclosed. A nuclear localization signal is used in fusion protein. Cell sorting, flow cytometry, microdissection, or microscopy observation, is used to sort cells. 11 ESTs and 6 unknown genes coding for proteins transporting into nucleus were identified by expressing as GFP fusion proteins in HepG2 cells and stimulating with TNFα. Eight unknown genes were also identified by heat shock stimulation.

RE.CNT 1 THERE ARE 1 CITED REFERENCES AVAILABLE FOR THIS RECORD
ALL CITATIONS AVAILABLE IN THE RE FORMAT

AB A method for screening a gene encoding a protein specifically transporting into nucleus by cloning each gene in a group of a large number of genes into an expression vector to **express** as **fusion protein** with a marker or **reporter** protein to construct a gene expression **library**, introducing this gene expression **library** into two cell groups to express the fusion proteins, stimulating one of the cell groups, separating cells wherein the fusion proteins are localized in the nucleus of such cells from both cell groups, and comparing the separated cells or nuclei to thereby search for a gene encoding a protein specifically transporting into the nucleus upon stimulus, is disclosed. A nuclear localization signal is used in fusion protein. Cell sorting, flow cytometry, microdissection, or microscopy observation, is used to sort cells. 11 ESTs and 6 unknown genes coding for proteins transporting into nucleus were identified by expressing as GFP fusion proteins in HepG2 cells and stimulating with TNFα. Eight unknown genes were also identified by heat shock stimulation.

IT Nucleic acid **library**
(gene expression; screening genes encoding nuclear transport proteins by GFP fusion protein expression and cell sorting)

L8 ANSWER 3 OF 4 CAPLUS COPYRIGHT 2004 ACS on STN
AN 1997:740333 CAPLUS
DN 128:10873
TI A three-hybrid reporter gene method for screening for proteins binding defined ligands

IN Liu, Jun; Licitra, Edward J.
PA Massachusetts Institute of Technology, USA
SO PCT Int. Appl., 40 pp.
CODEN: PIXXD2
DT Patent
LA English
FAN.CNT 1

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	WO 9741255	A1	19971106	WO 1997-US6912	19970425
	W: CA, JP				
	RW: AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE				
	CA 2252886	AA	19971106	CA 1997-2252886	19970425
	EP 907750	A1	19990414	EP 1997-921370	19970425
	EP 907750	B1	20020918		
	R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, FI				
	US 5928868	A	19990727	US 1997-845674	19970425
	JP 2000508923	T2	20000718	JP 1997-539036	19970425
	DE 29724617	U1	20020718	DE 1997-29724617	19970425
	EP 1241265	A2	20020918	EP 2002-76267	19970425
	EP 1241265	A3	20040218		
	R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, SI, LT, LV, FI, RO, MK, CY, AL				
	AT 224454	E	20021015	AT 1997-921370	19970425
	ES 2183166	T3	20030316	ES 1997-921370	19970425
PRAI	US 1996-17341P	P	19960426		
	EP 1997-921370	A3	19970425		
	WO 1997-US6912	W	19970425		

AB A method for identifying the binding partner for a define ligand using an extension of the two-hybrid system is described. The method uses a fusion protein of the LexA protein and a ligand binding protein to bind to a LexA operator upstream of a reporter gene. This is bound to by a conjugate of the natural ligand for the protein and the ligand of interest. Possible binding partners for the ligand are identified by introduction of an expression **library** in which the proteins are synthesized as fusion products with a transcriptional activator. When the necessary combination of LexA fusion protein, ligand, and transcriptional activator **fusion protein** are brought together, the **reporter** gene is **expressed**. The method is particularly intended for the identification of natural binding partners for small mols. A fusion product of LexA and the rat glucocorticoid receptor is used in a reconstruction experiment with FK506-binding protein FKBP12 is used to demonstrate using a conjugate of dexamethasone and FK506 as the hybrid ligand.
AB A method for identifying the binding partner for a define ligand using an extension of the two-hybrid system is described. The method uses a fusion protein of the LexA protein and a ligand binding protein to bind to a LexA operator upstream of a reporter gene. This is bound to by a conjugate of the natural ligand for the protein and the ligand of interest. Possible binding partners for the ligand are identified by introduction of an expression **library** in which the proteins are synthesized as fusion products with a transcriptional activator. When the necessary combination of LexA fusion protein, ligand, and transcriptional activator **fusion protein** are brought together, the **reporter** gene is **expressed**. The method is particularly intended for the identification of natural binding partners for small mols. A fusion product of LexA and the rat glucocorticoid receptor is used in a reconstruction experiment with FK506-binding protein FKBP12 is used to demonstrate using a conjugate of dexamethasone and FK506 as the hybrid ligand.

IT Combinatorial **library**
(nucleic acid, screening for ligand-binding proteins of; three-hybrid reporter gene method for screening for proteins binding defined

ligands)
IT Nucleic acid **library**
(screening for ligand-binding proteins of; three-hybrid reporter gene method for screening for proteins binding defined ligands)

L8 ANSWER 4 OF 4 MEDLINE on STN
AN 97140325 MEDLINE
DN PubMed ID: 8986806
TI A novel member of the RING finger family, KRIP-1, associates with the KRAB-A transcriptional repressor domain of zinc finger proteins.
AU Kim S S; Chen Y M; O'Leary E; Witzgall R; Vidal M; Bonventre J V
CS Renal Unit, Massachusetts General Hospital, Charlestown 02129, USA.
NC DK 38452 (NIDDK)
DK 39773 (NIDDK)
NS 10828 (NINDS)
SO Proceedings of the National Academy of Sciences of the United States of America, (1996 Dec 24) 93 (26) 15299-304.
Journal code: 7505876. ISSN: 0027-8424.
CY United States
DT Journal; Article; (JOURNAL ARTICLE)
LA English
FS Priority Journals
OS GENBANK-U67303
EM 199701
ED Entered STN: 19970219
Last Updated on STN: 19970219
Entered Medline: 19970128
AB The Kruppel-associated box A (KRAB-A) domain is an evolutionarily conserved transcriptional repressor domain present in approximately one-third of zinc finger proteins of the Cys2-His2 type. Using the yeast two-hybrid system, we report the isolation of a cDNA encoding a novel murine protein, KRAB-A interacting protein 1 (KRIP-1) that physically interacts with the KRAB-A region. KRIP-1 is a member of the RBCC subfamily of the RING finger, or Cys3HisCys4, family of zinc binding proteins whose other members are known to play important roles in differentiation, oncogenesis, and signal transduction. The KRIP-1 protein has high homology to TIF1, a putative modulator of ligand-dependent activation function of nuclear receptors. A 3.5-kb mRNA for KRIP-1 is ubiquitously expressed among all adult mouse tissues studied. When a GAL4-KRIP-1 **fusion protein** is **expressed** in COS cells with a chloramphenicol acetyltransferase **reporter** construct with five GAL4 binding sites, there is dose-dependent repression of transcription. Thus, KRIP-1 interacts with the KRAB-A region of C2H2 zinc finger proteins and may mediate or modulate KRAB-A transcriptional repressor activity.
AB . . . of nuclear receptors. A 3.5-kb mRNA for KRIP-1 is ubiquitously expressed among all adult mouse tissues studied. When a GAL4-KRIP-1 **fusion protein** is **expressed** in COS cells with a chloramphenicol acetyltransferase **reporter** construct with five GAL4 binding sites, there is dose-dependent repression of transcription. Thus, KRIP-1 interacts with the KRAB-A region of. . .
CT . . . Check Tags: Support, Non-U.S. Gov't; Support, U.S. Gov't, P.H.S.
Amino Acid Sequence
Animals
Base Sequence
COS Cells
Conserved Sequence
Embryo
Gene Library
Kidney: ME, metabolism
Mice
Molecular Sequence Data
Nuclear Proteins: BI, biosynthesis
Nuclear Proteins: CH, chemistry

*Nuclear Proteins: ME,. . .

```
=> s fusion(10a)green fluorescence(10a)librar###
L9            0 FUSION(10A) GREEN FLUORESCENCE(10A) LIBRAR###

=> s fusion(10a)green(10a)protein(10a)librar###
L10          40 FUSION(10A) GREEN(10A) PROTEIN(10A) LIBRAR###

=> s l10 and expres###
L11          15 L10 AND EXPRES###

=> s l11 and cycloheximide
L12           0 L11 AND CYCLOHEXIMIDE

=> l11 and screen###
L11 IS NOT A RECOGNIZED COMMAND
The previous command name entered was not recognized by the system.
For a list of commands available to you in the current file, enter
"HELP COMMANDS" at an arrow prompt (=>).

=> s l11 and screen###
L13          10 L11 AND SCREEN###

=> s l13 and (inhibit###(10a)degradat###)
L14           0 L13 AND (INHIBIT###(10A) DEGRADAT###)

=> s l13 and inhibit###
L15           0 L13 AND INHIBIT###

=> dup rem l13
PROCESSING COMPLETED FOR L13
L16           9 DUP REM L13 (1 DUPLICATE REMOVED)

=> d l16 1-9 bib ab kwic

L16  ANSWER 1 OF 9  CAPLUS  COPYRIGHT 2004 ACS on STN  DUPLICATE 1
AN   2003:296076  CAPLUS
DN   138:315791
TI   Fusions of random peptide libraries in scaffold proteins such as green
     fluorescent protein or β-lactamase
IN   Anderson, David; Peelle, Beau Robert; Bogenberger, Jakob Maria
PA   Rigel Pharmaceuticals, Inc., USA
SO   U.S., 63 pp., Cont.-in-part of U.S. 6,180,343.
     CODEN: USXXAM
DT   Patent
LA   English
FAN.CNT 4
     PATENT NO.         KIND  DATE          APPLICATION NO.  DATE
     ---------------    ----  --------      ---------------  --------
PI   US 6548632          B1   20030415      US 1999-415765   19991008
     US 6180343          B1   20010130      US 1998-169015   19981008
     US 6548249          B1   20030415      US 2000-626581   20000727
     US 6562617          B1   20030513      US 2000-626580   20000727
     US 2001003650       A1   20010614      US 2000-749959   20001227
     US 6596485          B2   20030722
     US 2003143562       A1   20030731      US 2002-177725   20020620
     US 2003224412       A1   20031204      US 2003-393449   20030318
PRAI US 1998-169015      A2   19981008
     US 1999-415765      A3   19991008
     US 2002-177725      A2   20020620
AB   The invention relates to the use of scaffold proteins, particularly
     green fluorescent protein (GFP) and β-lactamase
     TEM-1, in fusion constructs with random and defined peptides and
```

peptide **libraries**. The fusions act to increase the cellular expression levels, decrease the cellular catabolism, increase the conformational stability relative to linear peptides, and increase the steady state concns. of the random peptides and random peptide library members **expressed** in cells for the purpose of detecting the presence of the peptides and **screening** random peptide libraries. N-terminal, C-terminal, dual N- and C-terminal, and one or more internal fusions are all contemplated. Internal fusions in Renilla GFP may be made in loops 1-5 (amino acid residues 130-135, 154-159, 172-175, 188-193, or 208-216) for optimal presentation of the peptide. Inclusion of multiple highly flexible amino acid residues between GFP and the library allows minimal conformational constraints on the GFP. Designed insertion sites in loops 2-4 retain a high level of GFP fluorescence when the inserts are flanked by multiple glycines in the tetrapeptide linkers. Novel fusions utilizing self-binding peptides to create a conformationally stabilized fusion domain are also contemplated.

RE.CNT 44 THERE ARE 44 CITED REFERENCES AVAILABLE FOR THIS RECORD
ALL CITATIONS AVAILABLE IN THE RE FORMAT

AB The invention relates to the use of scaffold proteins, particularly **green** fluorescent **protein** (GFP) and β-lactamase TEM-1, in **fusion** constructs with random and defined peptides and peptide **libraries**. The fusions act to increase the cellular expression levels, decrease the cellular catabolism, increase the conformational stability relative to linear peptides, and increase the steady state concns. of the random peptides and random peptide library members **expressed** in cells for the purpose of detecting the presence of the peptides and **screening** random peptide libraries. N-terminal, C-terminal, dual N- and C-terminal, and one or more internal fusions are all contemplated. Internal fusions in Renilla GFP may be made in loops 1-5 (amino acid residues 130-135, 154-159, 172-175, 188-193, or 208-216) for optimal presentation of the peptide. Inclusion of multiple highly flexible amino acid residues between GFP and the library allows minimal conformational constraints on the GFP. Designed insertion sites in loops 2-4 retain a high level of GFP fluorescence when the inserts are flanked by multiple glycines in the tetrapeptide linkers. Novel fusions utilizing self-binding peptides to create a conformationally stabilized fusion domain are also contemplated.

ST scaffold protein fusion random peptide library; **green** fluorescent **protein fusion** random peptide **library**; lactamase fusion random peptide library

IT Animal cell line
(293, GFP fusions **expressed** in; fusions of random peptide libraries in scaffold proteins such as green fluorescent protein or β-lactamase)

IT Animal cell line
(JURKAT, GFP fusions **expressed** in; fusions of random peptide libraries in scaffold proteins such as green fluorescent protein or β-lactamase)

L16 ANSWER 2 OF 9 CAPLUS COPYRIGHT 2004 ACS on STN
AN 2003:950623 CAPLUS
DN 140:13712
TI Structurally biased random peptide libraries based on different scaffolds
IN Anderson, David; Peelle, Beau Robert; Bogenberger, Jakob Maria
PA USA
SO U.S. Pat. Appl. Publ., 110 pp., Cont.-in-part of U.S. Ser. No. 177,725.
CODEN: USXXCO
DT Patent
LA English
FAN.CNT 4

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	US 2003224412	A1	20031204	US 2003-393449	20030318
	US 6180343	B1	20010130	US 1998-169015	19981008

	US 6548632	B1	20030415	US 1999-415765	19991008
	US 2003143562	A1	20030731	US 2002-177725	20020620
PRAI	US 1998-169015	A2	19981008		
	US 1999-415765	A2	19991008		
	US 2002-177725	A2	20020620		

AB The invention relates to the use of scaffold proteins, particularly **green** fluorescent **protein** (GFP), in **fusion** constructs with random and defined peptides and peptide **libraries**, to increase the cellular expression levels, decrease the cellular catabolism, increase the conformational stability relative to linear peptides, and to increase the steady state concns. of the library peptides and peptide library members **expressed** in cells for the purpose of detecting the presence of the peptides and **screening** peptide libraries. N-terminal, C-terminal, dual N- and C-terminal and one or more internal fusions are all contemplated. Novel fusions utilizing self-binding peptides to create a conformationally stabilized fusion domain are also contemplated.

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ST scaffold protein fusion random peptide library; **green** fluorescent **protein fusion** random peptide **library**; lactamase fusion random peptide library

L16 ANSWER 3 OF 9 CAPLUS COPYRIGHT 2004 ACS on STN
AN 2003:590695 CAPLUS
DN 139:144918
TI Random peptide libraries in the context of a reporter protein scaffold for use in analysis of peptide effects on protein structure and function
IN Anderson, David; Peelle, Beau Robert; Bogenberger, Jakob Maria
PA Rigel Pharmaceuticals, Inc., USA
SO U.S. Pat. Appl. Publ., 110 pp., Cont.-in-part of U. S. Ser. No. 415,765.
CODEN: USXXCO
DT Patent
LA English
FAN.CNT 4

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	US 2003143562	A1	20030731	US 2002-177725	20020620
	US 6180343	B1	20010130	US 1998-169015	19981008
	US 6548632	B1	20030415	US 1999-415765	19991008
	US 2003224412	A1	20031204	US 2003-393449	20030318
PRAI	US 1998-169015	A2	19981008		
	US 1999-415765	A2	19991008		
	US 2002-177725	A2	20020620		

AB A method of using a reporter protein as a framework within which the effects of random peptides from a library on protein structure and function is described. A DNA library in which the gene for the reporter protein has sequences encoding random peptides inserted at defined sites is prepared and **expressed** in a suitable host. The library may be biased, e.g. to increase the possibility of including a peptide promoting an α-helical structure. The peptides may be at the N-terminal, the C-terminal, or at internal sites. The library is **screened** for members showing properties such as increases in cellular reporter concentration,

increased stability from lower rates of cellular catabolism, increased conformational stability relative to linear peptides, and increased steady state concns. of the library peptides. A reporter protein may be replaced by a protein essential for cell survival. Novel fusions utilizing self-binding peptides to create a conformationally stabilized fusion domain are also contemplated. Loops in green fluorescent were identified as potential sites for the peptide library and test peptides. Tests with two different insertions showed consistent effects on protein fluorescence in Escherichia coli and mammalian cells.

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increased stability from lower rates of cellular catabolism, increased conformational stability relative to linear peptides, and increased steady state concns. of the library peptides. A reporter protein may be replaced by a protein essential for cell survival. Novel fusions utilizing self-binding peptides to create a conformationally stabilized fusion domain are also contemplated. Loops in green fluorescent were identified as potential sites for the peptide library and test peptides. Tests with two different insertions showed consistent effects on protein fluorescence in Escherichia coli and mammalian cells.

ST scaffold protein fusion random peptide library; **green** fluorescent **protein fusion** random peptide **library**; lactamase fusion random peptide library

IT Peptides, biological studies
RL: BSU (Biological study, unclassified); BIOL (Biological study) (bioactive, **screening** for; random peptide libraries in context of reporter protein scaffold for use in anal. of peptide effects on protein structure and function)

L16 ANSWER 4 OF 9 BIOSIS COPYRIGHT 2004 BIOLOGICAL ABSTRACTS INC. on STN
AN 2003:378382 BIOSIS
DN PREV200300378382
TI Green fluorescent protein fusions with random peptides.
AU Anderson, David [Inventor, Reprint Author]; Bogenberger, Jakob Maria [Inventor]
CS Menlo Park, CA, USA
ASSIGNEE: Rigel Pharmaceuticals, Inc.
PI US 6596485 July 22, 2003
SO Official Gazette of the United States Patent and Trademark Office Patents, (July 22 2003) Vol. 1272, No. 4. http://www.uspto.gov/web/menu/patdata.htm 1. e-file.
ISSN: 0098-1133 (ISSN print).
DT Patent
LA English
ED Entered STN: 13 Aug 2003
Last Updated on STN: 13 Aug 2003

AB The invention relates to the use of fluorescent proteins, particularly **green** fluorescent **protein** (GFP), in **fusion** constructs with random and defined peptides and peptide **libraries**, to increase the cellular expression levels, decrease the cellular catabolism, increase the conformational stability relative to linear peptides, and to increase the steady state concentrations of the random peptides and random peptide library members **expressed** in cells for the purpose of detecting the presence of the peptides and **screening** random peptide libraries. N-terminal, C-terminal, dual N- and C-terminal and one or more internal fusions are all contemplated.

Novel fusions utilizing self-binding peptides to create a conformationally stabilized fusion domain are also contemplated.

AB The invention relates to the use of fluorescent proteins, particularly **green** fluorescent **protein** (GFP), in **fusion** constructs with random and defined peptides and peptide **libraries**, to increase the cellular expression levels, decrease the cellular catabolism, increase the conformational stability relative to linear peptides, and to increase the steady state concentrations of the random peptides and random peptide library members **expressed** in cells for the purpose of detecting the presence of the peptides and **screening** random peptide libraries. N-terminal, C-terminal, dual N- and C-terminal and one or more internal fusions are all contemplated. Novel fusions. . .

IT Methods & Equipment
random peptide library **screening**: laboratory techniques

L16 ANSWER 5 OF 9 BIOSIS COPYRIGHT 2004 BIOLOGICAL ABSTRACTS INC. on STN
AN 2003:280434 BIOSIS
DN PREV200300280434
TI Fusions of scaffold proteins with random peptide libraries.
AU Anderson, David [Inventor, Reprint Author]; Peelle, Beau Robert [Inventor]; Bogenberger, Jakob Maria [Inventor]
CS ASSIGNEE: Rigel Pharmaceuticals, Inc.
PI US 6562617 May 13, 2003
SO Official Gazette of the United States Patent and Trademark Office Patents, (May 13 2003) Vol. 1270, No. 2. http://www.uspto.gov/web/menu/patdata.html . e-file.
ISSN: 0098-1133 (ISSN print).
DT Patent
LA English
ED Entered STN: 11 Jun 2003
Last Updated on STN: 11 Jun 2003

AB The invention relates to the use of scaffold proteins, particularly **green** fluorescent **protein** (GFP), in **fusion** constructs with random and defined peptides and peptide **libraries**, to increase the cellular expression levels, decrease the cellular catabolism, increase the conformational stability relative to linear peptides, and to increase the steady state concentrations of the random peptides and random peptide library members **expressed** in cells for the purpose of detecting the presence of the peptides and **screening** random peptide libraries. N-terminal, C-terminal, dual N- and C-terminal and one or more internal fusions are all contemplated. Novel fusions utilizing self-binding peptides to create a conformationally stabilized fusion domain are also contemplated.

AB The invention relates to the use of scaffold proteins, particularly **green** fluorescent **protein** (GFP), in **fusion** constructs with random and defined peptides and peptide **libraries**, to increase the cellular expression levels, decrease the cellular catabolism, increase the conformational stability relative to linear peptides, and to increase the steady state concentrations of the random peptides and random peptide library members **expressed** in cells for the purpose of detecting the presence of the peptides and **screening** random peptide libraries. N-terminal, C-terminal, dual N- and C-terminal and one or more internal fusions are all contemplated. Novel fusions. . .

L16 ANSWER 6 OF 9 BIOSIS COPYRIGHT 2004 BIOLOGICAL ABSTRACTS INC. on STN
AN 2003:227250 BIOSIS
DN PREV200300227250
TI Fusions of scaffold proteins with random peptide libraries.
AU Anderson, David [Inventor, Reprint Author]; Peelle, Beau Robert [Inventor]; Bogenberger, Jakob Maria [Inventor]
CS San Bruno, CA, USA
ASSIGNEE: Rigel Pharmaceuticals, Inc.

PI US 6548249 April 15, 2003
SO Official Gazette of the United States Patent and Trademark Office Patents, (Apr 15 2003) Vol. 1269, No. 3. http://www.uspto.gov/web/menu/patdata.html . e-file.
ISSN: 0098-1133 (ISSN print).
DT Patent
LA English
ED Entered STN: 7 May 2003
Last Updated on STN: 7 May 2003
AB The invention relates to the use of scaffold proteins, particularly **green** fluorescent **protein** (GFP), in **fusion** constructs with random and defined peptides and peptide **libraries**, to increase the cellular expression levels, decrease the cellular catabolism, increase the conformational stability relative to linear peptides, and to increase the steady state concentrations of the random peptides and random peptide library members **expressed** in cells for the purpose of detecting the presence of the peptides and **screening** random peptide libraries. N-terminal, C-terminal, dual N- and C-terminal and one or more internal fusions are all contemplated. Novel fusions utilizing self-binding peptides to create a conformationally stabilized fusion domain are also contemplated.
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IT Methods & Equipment
peptide detection: laboratory techniques; random peptide library **screening**: laboratory techniques

L16 ANSWER 7 OF 9 CAPLUS COPYRIGHT 2004 ACS on STN
AN 2000:241464 CAPLUS
DN 132:261918
TI Fusions of scaffold proteins with random peptide libraries
IN Anderson, David; Bogenberger, Jakob Maria; Peelle, Beau Robert
PA Rigel Pharmaceuticals, Inc., USA
SO PCT Int. Appl., 89 pp.
CODEN: PIXXD2
DT Patent
LA English
FAN.CNT 4

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	WO 2000020574	A2	20000413	WO 1999-US23715	19991008
	WO 2000020574	A3	20000921		
	W: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, UZ, VN, YU, ZA, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM				
	RW: GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG				
	US 6180343	B1	20010130	US 1998-169015	19981008
	CA 2345215	AA	20000413	CA 1999-2345215	19991008
	EP 1119617	A2	20010801	EP 1999-957466	19991008
	R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT,				

IE, SI, LT, LV, FI, RO

	JP 2002526108	T2	20020820	JP 2000-574670	19991008
	AU 768126	B2	20031204	AU 2000-15164	19991008
	US 2001003650	A1	20010614	US 2000-749959	20001227
	US 6596485	B2	20030722		
PRAI	US 1998-169015	A	19981008		
	WO 1999-US23715	W	19991008		

AB The invention relates to the use of scaffold proteins, particularly **green** fluorescent **protein** (GFP), in **fusion** constructs with random and defined peptides and peptide **libraries**, to increase the cellular expression levels, decrease the cellular catabolism, increase the conformational stability relative to linear peptides, and to increase the steady state concns. of the random peptides and random peptide library members **expressed** in cells for the purpose of detecting the presence of the peptides and **screening** random peptide libraries. N-terminal, C-terminal, dual N- and C- terminal and one or more internal fusions are all contemplated. Novel fusions utilizing self-binding peptides to create a conformationally stabilized fusion domain are also contemplated. Thus, expts. to identify sites which may be used to insert peptides into GFP are described.

AB The invention relates to the use of scaffold proteins, particularly **green** fluorescent **protein** (GFP), in **fusion** constructs with random and defined peptides and peptide **libraries**, to increase the cellular expression levels, decrease the cellular catabolism, increase the conformational stability relative to linear peptides, and to increase the steady state concns. of the random peptides and random peptide library members **expressed** in cells for the purpose of detecting the presence of the peptides and **screening** random peptide libraries. N-terminal, C-terminal, dual N- and C- terminal and one or more internal fusions are all contemplated. Novel fusions utilizing self-binding peptides to create a conformationally stabilized fusion domain are also contemplated. Thus, expts. to identify sites which may be used to insert peptides into GFP are described.

ST scaffold protein peptide fusion library; **green** fluorescent **protein** peptide **fusion library**

L16 ANSWER 8 OF 9 CAPLUS COPYRIGHT 2004 ACS on STN
AN 2000:114405 CAPLUS
DN 132:147593
TI Methods and compositions for peptide libraries displayed on light-emitting scaffolds of proteins
IN Kamb, Carl Alexander; Abedi, Majid
PA Arcaris, Inc., USA
SO U.S., 24 pp., Cont.-in-part of U.S. 5,955,275.
CODEN: USXXAM
DT Patent
LA English
FAN.CNT 11

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	US 6025485	A	20000215	US 1997-965477	19971106
	US 6623922	B1	20030923	US 1997-800664	19970214
	US 5955275	A	19990921	US 1997-812994	19970304
	CA 2309543	AA	19990520	CA 1998-2309543	19981106
	WO 9924617	A1	19990520	WO 1998-US23778	19981106
	W: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM				
	RW: GH, GM, KE, LS, MW, SD, SZ, UG, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG				
	AU 9915200	A1	19990531	AU 1999-15200	19981106

EP 1029081 A1 20000823 EP 1998-959391 19981106
R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, FI
JP 2001522587 T2 20011120 JP 2000-519609 19981106
US 6566057 B1 20030520 US 1999-269006 19990317
NO 2000002375 A 20000630 NO 2000-2375 20000505
US 2002132229 A1 20020919 US 2001-929663 20010814
US 2002090605 A1 20020711 US 2001-935929 20010823
PRAI US 1997-800664 A2 19970214
US 1997-812994 A2 19970304
US 1996-699266 A2 19960819
WO 1997-US14514 A2 19970819
US 1997-965477 A 19971106
WO 1998-US23778 W 19981106
US 1999-259155 B1 19990226
US 1999-378420 B1 19990820

AB Methods and compns. for peptides or protein fragments displayed on scaffolds and libraries of sequences encoding peptides or protein fragments displayed on scaffolds that permit the properties of the library to be easily and quant. monitored are disclosed. The scaffold is a protein that is capable of emitting light. Thus, solvent-exposed loops are identified in the Aequorea victoria green fluorescent protein (GFP) which can accommodate and present random aptamers while allowing GFP to retain its autofluorescent properties; these loops optimally comprise amino acid residues Ala155-Ile161, Lys162-Gln183, and Gln184-Ser205. A minibody (Ig)-GFP scaffold system is also described. The plasmid vector pVT21, which permits induction of GFP expression in the presence of galactose, was obtained by manipulation of pACA151, a 6.7-kb 2μ yeast shuttle vector containing a red-shifted (S65T) GFP expression cassette and the phosphoglycerate kinase 3' end. Thus, anal. of the expression of individual members of the library when they are **expressed** in cells may be carried out using instruments that can analyze the emitted light, such as flow sorter (FACS), a spectrophotometer, a microtiter plate reader, a CCD, a fluorescence microscope, or other similar device. This permits **screening** of the expression library in host cells on a cell-by-cell basis, and enrichment of the library for sequences that have predetd. characteristics.

RE.CNT 7 THERE ARE 7 CITED REFERENCES AVAILABLE FOR THIS RECORD
ALL CITATIONS AVAILABLE IN THE RE FORMAT

AB Methods and compns. for peptides or protein fragments displayed on scaffolds and libraries of sequences encoding peptides or protein fragments displayed on scaffolds that permit the properties of the library to be easily and quant. monitored are disclosed. The scaffold is a protein that is capable of emitting light. Thus, solvent-exposed loops are identified in the Aequorea victoria green fluorescent protein (GFP) which can accommodate and present random aptamers while allowing GFP to retain its autofluorescent properties; these loops optimally comprise amino acid residues Ala155-Ile161, Lys162-Gln183, and Gln184-Ser205. A minibody (Ig)-GFP scaffold system is also described. The plasmid vector pVT21, which permits induction of GFP expression in the presence of galactose, was obtained by manipulation of pACA151, a 6.7-kb 2μ yeast shuttle vector containing a red-shifted (S65T) GFP expression cassette and the phosphoglycerate kinase 3' end. Thus, anal. of the expression of individual members of the library when they are **expressed** in cells may be carried out using instruments that can analyze the emitted light, such as flow sorter (FACS), a spectrophotometer, a microtiter plate reader, a CCD, a fluorescence microscope, or other similar device. This permits **screening** of the expression library in host cells on a cell-by-cell basis, and enrichment of the library for sequences that have predetd. characteristics.

IT Immunoglobulins
RL: ARU (Analytical role, unclassified); BPN (Biosynthetic preparation); BPR (Biological process); BSU (Biological study, unclassified); ANST (Analytical study); BIOL (Biological study); PREP (Preparation); PROC

(Process)
(**fusion** products, **green** fluorescent **protein** **fusion library**; methods and compns. for peptide **libraries** displayed on light-emitting scaffolds of proteins)

L16 ANSWER 9 OF 9 CAPLUS COPYRIGHT 2004 ACS on STN
AN 1999:330461 CAPLUS
DN 130:333721
TI Methods and compositions for peptide libraries displayed on light-emitting scaffolds of proteins
IN Kamb, Carl Alexander; Abedi, Majid
PA Ventana Genetics, Inc., USA
SO PCT Int. Appl., 56 pp.
CODEN: PIXXD2
DT Patent
LA English
FAN.CNT 11

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	WO 9924617	A1	19990520	WO 1998-US23778	19981106
	W: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM				
	RW: GH, GM, KE, LS, MW, SD, SZ, UG, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG				
	US 6025485	A	20000215	US 1997-965477	19971106
	CA 2309543	AA	19990520	CA 1998-2309543	19981106
	AU 9915200	A1	19990531	AU 1999-15200	19981106
	EP 1029081	A1	20000823	EP 1998-959391	19981106
	R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, FI				
	JP 2001522587	T2	20011120	JP 2000-519609	19981106
	US 6566057	B1	20030520	US 1999-269006	19990317
	NO 2000002375	A	20000630	NO 2000-2375	20000505
PRAI	US 1997-965477	A	19971106		
	US 1997-800664	A2	19970214		
	US 1997-812994	A2	19970304		
	WO 1998-US23778	W	19981106		

AB Methods and compns. for peptides or protein fragments displayed on scaffolds and libraries of sequences encoding peptides or protein fragments displayed on scaffolds that permit the properties of the library to be easily and quant. monitored are disclosed. The scaffold is a protein that is capable of emitting light. Thus, anal. of the expression of individual members of the library when they are **expressed** in cells may be carried out using instruments that can analyze the emitted light, such as a flow sorter (FACS), a spectrophotometer, a microtiter plate reader, a CCD, a fluorescence microscope, or other similar device. This permits **screening** of the expression library in host cells on a cell-by-cell basis, and enrichment of the library for sequences that have predetd. characteristics.
RE.CNT 4 THERE ARE 4 CITED REFERENCES AVAILABLE FOR THIS RECORD
ALL CITATIONS AVAILABLE IN THE RE FORMAT
AB Methods and compns. for peptides or protein fragments displayed on scaffolds and libraries of sequences encoding peptides or protein fragments displayed on scaffolds that permit the properties of the library to be easily and quant. monitored are disclosed. The scaffold is a protein that is capable of emitting light. Thus, anal. of the expression of individual members of the library when they are **expressed** in cells may be carried out using instruments that can analyze the emitted light, such as a flow sorter (FACS), a spectrophotometer, a microtiter plate reader, a CCD, a fluorescence microscope, or other similar device.

This permits **screening** of the expression library in host cells on a cell-by-cell basis, and enrichment of the library for sequences that have predetd. characteristics.

ST peptide **library fusion** light emitting scaffold **green** fluorescent **protein**

IT Immunoglobulins
RL: ARU (Analytical role, unclassified); BPN (Biosynthetic preparation); BPR (Biological process); BSU (Biological study, unclassified); ANST (Analytical study); BIOL (Biological study); PREP (Preparation); PROC (Process)
(**green** fluorescent **protein fusion library**; methods and compns. for peptide **libraries** displayed on light-emitting scaffolds of proteins)

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